Correction: Genome-wide physical activity interactions in adiposity — A meta-analysis of 200,452 adults

There are errors in some of the gene names in S7, S8, S9, S10, S11 and S12 Tables. Please view the correct tables below, with the locus numbers removed since they are not defined across all the models in the paper.

Supporting information

S7 Table. All SNPs that met significance for BMI in the European only analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S8 Table. All SNPs that met significance for BMI in the all ancestry analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S9 Table. All SNPs that met significance for waist circumference adjusted for BMI in the European only analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S10 Table. All SNPs that met significance for waist circumference adjusted for BMI in the all ancestry analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S11 Table. All SNPs that met significance for waist-to-hip ratio adjusted for BMI in the European only analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S12 Table. All SNPs that met significance for waist-to-hip ratio adjusted for BMI in the all ancestry analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)
Reference
RESEARCH ARTICLE

Genome-wide physical activity interactions in adiposity – A meta-analysis of 200,452 adults


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Abstract

Physical activity (PA) may modify the genetic effects that give rise to increased risk of obesity. To identify adiposity loci whose effects are modified by PA, we performed genome-wide interaction meta-analyses of BMI and BMI-adjusted waist circumference and waist-hip ratio from up to 200,452 adults of European (n = 180,423) or other ancestry (n = 20,029). We standardized PA by categorizing it into a dichotomous variable where, on average, 23% of participants were categorized as inactive and 77% as physically active. While we replicate the interaction with PA for the strongest known obesity-risk locus in the FTO gene, of which the effect is attenuated by ~30% in physically active individuals compared to inactive individuals, we do not identify additional loci that are sensitive to PA. In additional genome-wide meta-analyses adjusting for PA and interaction with PA, we identify 11 novel adiposity loci, suggesting that accounting for PA or other environmental factors that contribute to variation in adiposity may facilitate gene discovery.

Author summary

Decline in daily physical activity is thought to be a key contributor to the global obesity epidemic. However, the impact of sedentariness on adiposity may be in part determined by a person’s genetic constitution. The specific genetic variants that are sensitive to physical activity and regulate adiposity remain largely unknown. Here, we aimed to identify genetic variants whose effects on adiposity are modified by physical activity by examining ~2.5 million genetic variants in up to 200,452 individuals. We also tested whether adjusting for physical activity as a covariate could lead to the identification of novel adiposity variants. We find robust evidence of interaction with physical activity for the strongest known obesity risk-locus in the FTO gene, of which the body mass index-increasing effect is attenuated by ~30% in physically active individuals compared to inactive individuals. Our analyses indicate that other similar gene-physical activity interactions may exist, but better measurement of physical activity, larger sample sizes, and/or improved analytical methods will be required to identify them. Adjusting for physical activity, we identify 11 novel adiposity variants, suggesting that accounting for physical activity or other environmental factors that contribute to variation in adiposity may facilitate gene discovery.

Introduction

In recent decades, we have witnessed a global obesity epidemic that may be driven by changes in lifestyle such as easier access to energy-dense foods and decreased physical activity (PA) [1]. However, not everyone becomes obese in obesogenic environments. Twin studies suggest that changes in body weight in response to lifestyle interventions are in part determined by a person’s genetic constitution [2–4]. Nevertheless, the genes that are sensitive to environmental influences remain largely unknown.

Previous studies suggest that genetic susceptibility to obesity, assessed by a genetic risk score for BMI, may be attenuated by PA [5, 6]. A large-scale meta-analysis of the FTO obesity locus in 218,166 adults showed that being physically active attenuates the BMI increasing effect of this locus by ~30% [7]. While these findings suggest that FTO, and potentially other
previously established BMI loci, may interact with PA, it has been hypothesized that loci showing the strongest main effect associations in genome-wide association studies (GWAS) may be the least sensitive to environmental and lifestyle influences, and may therefore not make the best candidates for interactions [8]. Yet no genome-wide search for novel loci exhibiting SNP×PA interaction has been performed. A genome-wide meta-analysis of genotype-dependent phenotypic variance of BMI, a marker of sensitivity to environmental exposures, in ~170,000 participants identified FTO, but did not show robust evidence of environmental sensitivity for other loci [9]. Recent genome-wide meta-analyses of adiposity traits in >320,000 individuals uncovered loci interacting with age and sex, but also suggested that very large sample sizes are required for interaction studies to be successful [10].

Here, we report results from a large-scale genome-wide meta-analysis of SNP×PA interactions in adiposity in up to 200,452 adults. As part of these interaction analyses, we also examine whether adjusting for PA or jointly testing for SNP’s main effect and interaction with PA may identify novel adiposity loci.

Results

Identification of loci interacting with PA

We performed meta-analyses of results from 60 studies, including up to 180,423 adults of European descent and 20,029 adults of other ancestries to assess interactions between ~2.5 million genotyped or HapMap-imputed SNPs and PA on BMI and BMI-adjusted waist circumference (WC adjBMI) and waist-hip ratio (WHR adjBMI) (S1–S5 Tables). Similar to a previous meta-analysis of the interaction between FTO and PA [7], we standardized PA by categorizing it into a dichotomous variable where on average ~23% of participants were categorized as inactive and ~77% as physically active (see Methods and S6 Table). On average, inactive individuals had 0.99 kg/m^2 higher BMI, 3.46 cm higher WC, and 0.018 higher WHR than active individuals (S4 and S5 Tables).

Each study first performed genome-wide association analyses for each SNP’s effect on BMI in the inactive and active groups separately. Corresponding summary statistics from each cohort were subsequently meta-analyzed, and the SNP×PA interaction effect was estimated by calculating the difference in the SNP’s effect between the inactive and active groups. To identify sex-specific SNP×PA interactions, we performed the meta-analyses separately in men and women, as well as in the combined sample. In addition, we carried out meta-analyses in European-ancestry studies only and in European and other-ancestry studies combined.

We used two approaches to identify loci whose effects are modified by PA. In the first approach, we searched for genome-wide significant SNP×PA interaction effects (P INT < 5x10^{-8}). As shown in Fig 1, this approach yielded the highest power to identify cross-over interaction effects where the SNP’s effect is directionally opposite between the inactive and active groups. However, this approach has low power to identify interaction effects where the SNP’s effect is directionally concordant between the inactive and active groups (Fig 1). We identified a genome-wide significant interaction between rs986732 in cadherin 12 (CDH12) and PA on BMI in European-ancestry studies (beta INT = -0.076 SD/allele, P INT = 3.1x10^{-8}, n = 134,767) (S7 Table). The interaction effect was directionally consistent but did not replicate in an independent sample of 31,097 individuals (beta INT = -0.019 SD/allele, P INT = 0.52), and the pooled association P value for the discovery and replication stages combined did not reach genome-wide significance (N TOTAL = 165,864; P INT-TOTAL = 3x10^{-7}) (S1 Fig). No loci showed genome-wide significant interactions with PA on WC adjBMI or WHR adjBMI. CDH12 encodes an integral membrane protein mediating calcium-dependent cell-cell adhesion in the brain, where it may play a role in neurogenesis [11]. While CDH12 rs4701252 and rs268972 SNPs have shown suggestive
Fig 1. Power to identify PA-adjusted main, joint or GxPA interaction effects in 200,000 individuals (45,000 inactive, 155,000 active). The plots compare power to identify genome-wide significant main effects ($P_{\text{adjPA}} < 5 \times 10^{-8}$, dashed black), joint effects ($P_{\text{JOINT}} < 5 \times 10^{-5}$, dotted green) or GxPA interaction effects ($P_{\text{INT}} < 5 \times 10^{-8}$, solid magenta) as well as the power to identify Bonferroni-corrected interaction effects ($P_{\text{INT}} < 0.05/\text{number of loci}$, solid orange) for the SNPs that reached a genome-wide significant PA-adjusted main effect association ($P_{\text{adjPA}} < 5 \times 10^{-8}$). The power computations were based on analytical power formulae provided elsewhere [50] and were conducted a-priori based on various types of
known realistic BMI effect sizes [51]. Panels A, C, E: Assuming an effect in inactive individuals similar to a small ($R_{\text{INACT}} = 0.01\%$, comparable to the known BMI effect of the NUDT3 locus), medium ($R_{\text{INACT}} = 0.07\%$, comparable to the known BMI effect of the BDNF locus) and large ($R_{\text{INACT}} = 0.34\%$, comparable to the known BMI effect of the FTO locus) realistic effect on BMI and for various effects in physically active individuals (varied on the x axis); Panels B, D, F: Assuming an effect in physically active individuals similar to the small, medium and large realistic effects of the NUDT3, BDNF and FTO loci on BMI and for various effects in inactive individuals (varied on x axis).

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associations with waist circumference ($P = 2\times10^{-6}$) and BMI ($P = 5\times10^{-5}$) in previous GWAS [12, 13], the SNPs are not in LD with rs986732 ($r^2 < 0.1$).

In our second approach, we tested interaction for loci showing a genome-wide significant main effect on BMI, WC_{adjBMI} or WHR_{adjBMI} (S7–S12 Tables). We adjusted the significance threshold for SNP×PA interaction by Bonferroni correction ($P = 0.05/\text{number of SNPs tested}$).

As shown in Fig 1, this approach enhanced our power to identify interaction effects where there is a difference in the magnitude of the SNP’s effect between inactive and active groups when the SNP’s effect is directionally concordant between the groups. We identified a significant SNP×PA interaction of the FTO rs9941349 SNP on BMI in the meta-analysis of European-ancestry individuals; the BMI-increasing effect was 33% smaller in active individuals ($\text{beta}_{\text{ACTIVE}} = 0.072 \text{ SD/allele}$) than in inactive individuals ($\text{beta}_{\text{INACTIVE}} = 0.106 \text{ SD/allele}$, $P_{\text{INT}} = 4\times10^{-8}$). The rs9941349 SNP is in strong LD ($r^2 = 0.87$) with FTO rs9939609 for which interaction with PA has been previously established in a meta-analysis of 218,166 adults [7]. We identified no loci interacting with PA for WC_{adjBMI} or WHR_{adjBMI}.

In a previously published meta-analysis [7], the FTO locus showed a geographic difference for the interaction effect where the interaction was more pronounced in studies from North America than in those from Europe. To test for geographic differences in the present study, we performed additional meta-analyses for the FTO rs9941349 SNP on BMI in the meta-analysis of European-ancestry individuals; the interaction effect was more pronounced in studies from North America (beta_{INT} = 0.052 SD/allele, $P = 5\times10^{-4}$, $N = 63,896$) than in those from Europe (beta_{INT} = 0.028 SD/allele, $P = 0.006$, $N = 109,806$), we did not find a statistically significant difference between the regions ($P = 0.14$).

Explained phenotypic variance in inactive and active individuals. We tested whether the variance explained by $\sim 1.1$ million common variants (MAF $\geq 1\%$) differed between the inactive and active groups for BMI, WC_{adjBMI} and WHR_{adjBMI} [14]. In the physically active individuals, the variants explained $\sim 20\%$ less of variance in BMI than in inactive individuals (12.4% vs. 15.7%, respectively; $P_{\text{difference}} = 0.046$), suggesting that PA may reduce the impact of genetic predisposition to adiposity overall. There was no significant difference in the variance explained between active and inactive groups for WC_{adjBMI} (8.6% for active, 9.3% for inactive; $P_{\text{difference}} = 0.70$) or WHR_{adjBMI} (6.9% for active, 8.0% for inactive; $P_{\text{difference}} = 0.59$).

To further investigate differences in explained variance between the inactive and active groups, we calculated variance explained by subsets of SNPs selected based on significance thresholds (ranging from $P = 5\times10^{-8}$ to $P = 0.05$) of PA-adjusted SNP association with BMI, WC_{adjBMI} or WHR_{adjBMI} [15] (S13 Table). We found 17–26% smaller explained variance for BMI in the active group than in the inactive group at all P value thresholds (S13 Table).

Identification of novel loci when adjusting for PA or when jointly testing for SNP main effect and interaction with PA

Physical activity contributes to variation in BMI, WC_{adjBMI} and WHR_{adjBMI}, hence, adjusting for PA as a covariate may enhance power to identify novel adiposity loci. To that extent, each study performed genome-wide analyses for association with BMI, WC_{adjBMI} and WHR_{adjBMI} while adjusting for PA. Subsequently, we performed meta-analyses of the study-specific
results. We discovered 10 genome-wide significant loci (2 for BMI, 1 for \( WC_{adjBMI} \), 7 for \( WHR_{adjBMI} \)) that have not been reported in previous GWAS of adiposity traits (Table 1, S2–S4 Figs).

To establish whether additionally accounting for SNP×PA interactions would identify novel loci, we calculated the joint significance of PA-adjusted SNP main effect and SNP×PA interaction using the method of Aschard et al [16]. As illustrated in Fig 1, the joint test enhanced our power to identify loci where the SNP shows simultaneously a main effect and an interaction effect. We identified a novel BMI locus near \( ELAVL2 \) in men (\( P_{\text{JOINT}} = 4 \times 10^{-8} \)), which also showed suggestive evidence of interaction with PA (\( P_{\text{INT}} = 9 \times 10^{-4} \)); the effect of the BMI-increasing allele was attenuated by 71% in active as compared to inactive individuals (\( \beta_{\text{INACTIVE}} = 0.087 \text{SD/allele}, \beta_{\text{ACTIVE}} = 0.025 \text{SD/allele} \) (Table 1, S2–S4 Figs).

To evaluate the effect of PA adjustment on the results for the 11 novel loci, we performed a look-up in published GIANT consortium meta-analyses for BMI, \( WC_{adjBMI} \), and \( WHR_{adjBMI} \) that did not adjust for PA [17, 18] (S22 Table). All 11 loci showed a consistent direction of effect between the present PA-adjusted and the previously published PA-unadjusted results, but the PA-unadjusted associations were less pronounced despite up to 40% greater sample size, suggesting that adjustment for PA may have increased our power to identify these loci.

The biological relevance of putative candidate genes in the novel loci, based on our thorough searches of the literature, GWAS catalog look-ups, and analyses of eQTL enrichment and overlap with functional regulatory elements, are described in Tables 2 and 3. As the novel loci were identified in a PA-adjusted model, where adjusting for PA may have contributed to their identification, we examined whether the lead SNPs in these loci are associated with the level of PA. More specifically, we performed look-ups in GWAS analyses for the levels of moderate-to-vigorous intensity leisure-time PA (\( n = 80,035 \)), TV-viewing time (\( n = 28,752 \)), and sedentary behavior at work (\( n = 59,381 \)) or during transportation (\( n = 15,152 \)) [personal communication with Marcel den Hoed, Marilyn Cornelis, and Ruth Loos]. However, we did not find significant associations when correcting for the number of loci that were examined (\( P > 0.005 \)) (S16 Table).

Identification of secondary signals

In addition to uncovering 11 novel adiposity loci, our PA-adjusted GWAS and the joint test of SNP main effect and SNP×PA interaction confirmed 148 genome-wide significant loci (50 for BMI, 58 for \( WC_{adjBMI} \), 40 for \( WHR_{adjBMI} \)) that have been established in previous main effect GWAS for adiposity traits (S7–S12 Tables, S4 Fig). The lead SNPs in eight of the previously established loci (5 for BMI, 3 for \( WC_{adjBMI} \)), however, showed no LD or only weak LD \( (r^2 < 0.3) \) with the published lead SNP, suggesting they could represent novel secondary signals in known loci (S17 Table). To test whether these eight signals are independent of the previously published signals, we performed conditional analyses [19]. Three of the eight SNPs we examined, in/near \( NDUFS4 \), \( MEF2C-AS1 \) and \( CPA1 \), were associated with \( WC_{adjBMI} \) with \( P < 5 \times 10^{-8} \) in our PA-adjusted GWAS even after conditioning on the published lead SNP, hence representing novel secondary signals in these loci (S17 Table).

Enrichment of the identified loci with functional regulatory elements

Epigenetic variation may underlie gene-environment interactions observed in epidemiological studies [20] and PA has been shown to induce marked epigenetic changes in the genome [21]. We examined whether the BMI or \( WHR_{adjBMI} \) loci reaching \( P < 1 \times 10^{-5} \) for interaction with PA (13 loci for BMI, 5 for \( WHR_{adjBMI} \)) show overall enrichment with chromatin states in adipose, brain and muscle tissues available from the Roadmap Epigenomics Consortium [22].
Table 1. Novel loci achieving genome-wide significance (P<5x10^{-8}) in meta-analyses for PA-adjusted SNP main effect (P_{adjPA}) or the joint test of SNP main effect and SNP-PA interaction (P_{joint}).

<table>
<thead>
<tr>
<th>Trait</th>
<th>Marker</th>
<th>Nearest Gene</th>
<th>Chr</th>
<th>Pos (hg19)</th>
<th>Trait increasing/decreasing allele</th>
<th>Trait increasing allele's frequency</th>
<th>Analysis</th>
<th>N_{adjPA}</th>
<th>Beta_{adjPA}</th>
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<th>P_{adjPA}</th>
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<td>WHR_{adjBMI}</td>
<td>rs2300481</td>
<td>MEIS1</td>
<td>2</td>
<td>66,782,467</td>
<td>T/C</td>
<td>0.39</td>
<td>Overall</td>
<td>110881</td>
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<td>2.41E-08</td>
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<td>3.93E-08</td>
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<td>Women</td>
<td>66519</td>
<td>0.0288</td>
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<td>Men</td>
<td>43845</td>
<td>0.0258</td>
<td>0.0073</td>
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<td>rs167025</td>
<td>ARHGEF28</td>
<td>5</td>
<td>73,433,308</td>
<td>A/G</td>
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<td>70494</td>
<td>0.0223</td>
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<td>46591</td>
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<td>6</td>
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<td>G/A</td>
<td>0.87</td>
<td>Overall</td>
<td>149338</td>
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<tr>
<th>Trait Marker Nearest Gene</th>
<th>Chr</th>
<th>Pos (hg19)</th>
<th>Trait increasing/decreasing allele</th>
<th>Trait increasing allele’s frequency</th>
<th>Analysis</th>
<th>N_adjPA</th>
<th>Beta_adjPA</th>
<th>SE_adjPA</th>
<th>P_adjPA</th>
<th>P_int</th>
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<td>WHR_adjBMI rs6976930 BAZ1B</td>
<td>7</td>
<td>72,885,810</td>
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<td>Overall</td>
<td>145913</td>
<td>0.0294</td>
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<td>Women</td>
<td>83184</td>
<td>0.0326</td>
<td>0.0066</td>
<td>7.70E-07</td>
<td>7.00E-01</td>
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<td>62149</td>
<td>0.0254</td>
<td>0.0075</td>
<td>7.69E-04</td>
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<td>WHR_adjBMI rs10786152 PLCE1</td>
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<td>95,893,514</td>
<td>A/G</td>
<td>0.52</td>
<td>Overall</td>
<td>147123</td>
<td>0.0224</td>
<td>0.0051</td>
<td>1.79E-08</td>
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<td>1.44E-08</td>
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<td>Women</td>
<td>83884</td>
<td>0.0192</td>
<td>0.0051</td>
<td>1.56E-04</td>
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<td>62722</td>
<td>0.0255</td>
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<td>WHR_adjBMI rs889512 CTRB2</td>
<td>16</td>
<td>75,242,012</td>
<td>C/G</td>
<td>0.88</td>
<td>Overall</td>
<td>117417</td>
<td>0.031</td>
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<td>2.70E-05</td>
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<td>Women</td>
<td>70315</td>
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<td>2.67E-08</td>
<td>9.96E-02</td>
<td>1.09E-07</td>
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<td></td>
<td>Men</td>
<td>46440</td>
<td>-0.0022</td>
<td>0.0114</td>
<td>8.50E-01</td>
<td>5.06E-01</td>
<td>7.80E-01</td>
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</table>

**Novel loci achieving genome-wide significance in all-ancestry meta-analyses**

<table>
<thead>
<tr>
<th>Trait Marker Nearest Gene</th>
<th>Chr</th>
<th>Pos (hg19)</th>
<th>Trait increasing/decreasing allele</th>
<th>Trait increasing allele’s frequency</th>
<th>Analysis</th>
<th>N_adjPA</th>
<th>Beta_adjPA</th>
<th>SE_adjPA</th>
<th>P_adjPA</th>
<th>P_int</th>
<th>P_joint</th>
</tr>
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<tbody>
<tr>
<td>BMI rs754635 CCK</td>
<td>3</td>
<td>42,305,131</td>
<td>G/C</td>
<td>0.87</td>
<td>Overall</td>
<td>151282</td>
<td>0.0356</td>
<td>0.0062</td>
<td>1.07E-08</td>
<td>1.21E-01</td>
<td>3.28E-07</td>
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<td></td>
<td>Women</td>
<td>91241</td>
<td>0.026</td>
<td>0.0079</td>
<td>9.66E-04</td>
<td>1.25E-01</td>
<td>8.69E-04</td>
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<td></td>
<td>Men</td>
<td>62741</td>
<td>0.0486</td>
<td>0.0093</td>
<td>1.61E-07</td>
<td>2.98E-01</td>
<td>3.68E-06</td>
</tr>
</tbody>
</table>

Chr: chromosome; Pos(hg19): position based on human assembly 19; N_adjPA, Beta_adjPA, SE_adjPA, or P_adjPA: sample size, effect size, standard error, or P value, respectively, in the physical activity adjusted SNP main effect model; PA: physical activity; WC_adjBMI: BMI-adjusted waist circumference; WHR_adjBMI: BMI-adjusted waist-hip ratio; P_int: P value for SNP-PA interaction; P_joint: P value for the joint test of SNP main effect and SNP-PA interaction.

https://doi.org/10.1371/journal.pgen.1006528.t001
However, we did not find significant enrichment (S18 and S19 Tables), which may be due to the limited number of identified loci. The lack of significant findings may also be due to the assessment of chromatin states in the basal state, which may not reflect the dynamic changes that occur when cells are perturbed by PA [23].

We also tested whether the loci reaching $P < 5 \times 10^{-8}$ in our PA-adjusted GWAS of BMI or WHR$_{adjBMI}$ show enrichment with chromatin states and found significant enrichment of the BMI loci with enhancer, weak transcription, and polycomb-repressive elements in several brain cell lines, and with enhancer elements in three muscle cell lines (Table 2).

We also found significant enrichment of the WHR$_{adjBMI}$ loci with enhancer elements in three adipose and six muscle cell lines, with active transcription start sites in two adipose cell lines, and with polycomb-repressive elements in seven brain cell lines. The enrichment of our PA-adjusted main effect results with chromatin annotations in skeletal muscle in particular, the tissue most affected by PA, could highlight regulatory mechanisms that may be influenced by PA.

**Discussion**

In this genome-wide meta-analysis of more than 200,000 adults, we do not find evidence of interaction with PA for loci other than the established FTO locus. However, when adjusting for PA or jointly testing for SNP main effect and interaction with PA, we identify 11 novel adiposity loci, suggesting that accounting for PA or other environmental factors that contribute to variation in adiposity may increase power for gene discovery.

Our results suggest that if SNP×PA interaction effects for common variants exist, they are unlikely to be of greater magnitude than observed for FTO, the BMI-increasing effect of which is attenuated by ~30% in physically active individuals. The fact that common SNPs

<table>
<thead>
<tr>
<th>Table 2. Genes of biological interest within 500 kb of lead SNPs associated with BMI.</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>CCK (rs754635):</strong> The lead SNP is located in intron 1 of the CCK gene that encodes cholecystokinin, a gastrointestinal peptide that stimulates the digestion of fat and protein in the small intestine by inhibiting gastric emptying, inducing the release of pancreatic enzymes, increasing production of hepatic bile, and causing contraction of the gallbladder. Cholecystokinin induces satiety and reduces the amount of food consumed when administered prior to a meal [52, 53]. In a candidate gene study, four common variants in CCK were associated with increased meal size [54], but the variants are not in LD with rs754635 ($r^2 &lt; 0.1$). A GWAS of BMI in 62,246 individuals of East Asian ancestry showed a suggestive association ($P = 2 \times 10^{-7}$) for the rs4377469 SNP in high LD with our lead SNP ($r^2 = 0.7$) [55].</td>
</tr>
<tr>
<td><strong>ELAVL2 (rs1934100):</strong> The lead SNP showed an association with BMI only in men (Table 1). The only nearby gene ELAVL2 (455 kb away) is a conserved neuron-specific RNA-binding protein involved in stabilization or enhanced translation of specific mRNAs with AU-rich elements in the 3'-untranslated region [56]. While ELAVL2 is implicated in neuronal differentiation [56], potential mechanisms linking this function to obesity remain unclear.</td>
</tr>
<tr>
<td><strong>MRAS (rs1720825):</strong> The lead SNP is an intronic variant in MRAS. The MRAS rs1199333 SNP, in high LD with rs1720825 ($r^2 = 0.85$), has shown suggestive association with typical sporadic amyotrophic lateral sclerosis in a Chinese Han population ($P = 4 \times 10^{-6}$, S14 Table). Other MRAS SNPs have been associated with risk of coronary artery disease [57] but they are not in LD with rs1720825 ($r^2 &lt; 0.06$). MRAS encodes a member of the membrane-associated Ras small GTPase protein family that function as signal transducers in multiple processes of cell growth and differentiation and are involved in energy expenditure, adipogenesis, muscle differentiation, insulin signaling and glucose metabolism [58–60]. Mice with Mras knockout develop a severe obesity phenotype [61]. The SNP rs1199334, in high LD with our lead SNP rs1720825 ($r^2 = 0.90$), has been identified as the SNP most strongly associated with the cis-expression of centrosomal protein 70kDa (CEP70) in subcutaneous adipose tissue ($P = 2 \times 10^{-7}$) (S15 Table). CEP70 encodes a centrosomal protein that is critical for the regulation of mitotic spindle assembly, playing an essential role in cell cycle progression [62].</td>
</tr>
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</table>

https://doi.org/10.1371/journal.pgen.1006528.t002

However, we did not find significant enrichment (S18 and S19 Tables), which may be due to the limited number of identified loci. The lack of significant findings may also be due to the assessment of chromatin states in the basal state, which may not reflect the dynamic changes that occur when cells are perturbed by PA [23].
Table 3. Genes of biological interest within 500 kb of lead SNPs associated with WC_{adjBMI} or WHR_{adjBMI}.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Description</th>
<th>LD with</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>ZSCAN2 (rs7176527):</strong></td>
<td>Twenty two genes lie within 500kb of the WC_{adjBMI}-associated lead SNP (S3 Fig). The nearest gene, ZSCAN2, contains several copies of a zinc finger motif commonly found in transcriptional regulatory proteins. The rs7176527 SNP is in LD (r^2&gt;0.8) with five SNPs (rs3762168, rs2762169, rs12594450, rs72630460, and rs16974951) that are enhancers in multiple tissues in the data from Roadmap Epigenomics Consortium [22]. The rs7176527 SNP is cis-eQTL for the putative transcriptional regulator SCAND2 [63] in the intestine, prefrontal cortex, and lymphocytes (S15 Table).</td>
<td><strong>0.8</strong></td>
<td>[22, 63]</td>
</tr>
<tr>
<td><strong>PAPPA2 (rs4650943):</strong></td>
<td>Seven genes lie within 500kb of the lead SNP (S3 Fig). The nearest gene, PAPPA2, is 18 kb upstream of rs4650943 and codes for a protease that locally regulates insulin-like growth factor availability through cleavage of IGF binding protein 5, most commonly found in bone tissue. In murine models, the PAPP-A2 protein has been shown to influence overall body size and bone growth, but not glucose metabolism or adiposity [64–66].</td>
<td><strong>0.8</strong></td>
<td>[64–66]</td>
</tr>
<tr>
<td><strong>MEIS1 (rs2300481):</strong></td>
<td>The only gene within 500 kb of the lead SNP is MEIS1 encoding a homeobox protein that plays an important role in normal organismal growth and development. Two variants in high LD with the lead SNP (r^2 = 0.95) have been identified for association with PR interval of the heart (S14 Table). Another variant, in low LD with rs2300481 (r^2 = 0.25), has been associated with restless leg syndrome [67]–a sleeping disorder that may cause weight gain [68].</td>
<td><strong>0.8</strong></td>
<td>[67, 68]</td>
</tr>
<tr>
<td><strong>ARHGEF28 (rs167025):</strong></td>
<td>The lead SNP showed an association with WHR_{adjBMI} in men only (Table 1). There are two protein-coding genes within 500kb of rs167025. The nearest gene is ARHGEF28, 195 kb downstream, encoding Rho guanine nucleotide exchange factor 28. This exchange factor has been shown to destabilize low molecular weight neurofilament mRNAs in patients with amyotrophic lateral sclerosis, leading to degeneration and death of motor neurons controlling voluntary muscle movement [69, 70]. The ENC1 gene, 490 kb away, encodes Ectoderm-neural cortex protein 1, an actin-binding protein required for adipocyte differentiation [71].</td>
<td><strong>0.8</strong></td>
<td>[69, 70, 71]</td>
</tr>
<tr>
<td><strong>HCP5 (rs3094013):</strong></td>
<td>The lead SNP showed an association with WHR_{adjBMI} in men only (Table 1). The rs3094013 SNP is located in the MHC complex on chromosome 6, and the region within 500kb contains 124 genes (S3 Fig). The known WHR_{adjBMI}-increasing allele rs3099844, in strong LD with our lead SNP (r^2 &gt; 0.8), has previously been associated with increased HDL-cholesterol levels [72]. Candidate gene studies suggest that rs1800629 in tumor necrosis factor (TNF), which is 109 kb upstream and in moderate LD (r^2 = 0.64) with the lead SNP, may interact with physical activity to decrease serum CRP levels [73, 74]. We did not, however, find an interaction between rs1800629 and physical activity on WHR_{adjBMI} (P = 0.3).</td>
<td><strong>0.8</strong></td>
<td>[72, 73, 74]</td>
</tr>
<tr>
<td><strong>BAZ1B (rs6976930):</strong></td>
<td>There are 31 genes within 500kb the lead SNP rs6976930 (S3 Fig) which is in high LD (r^2&gt;0.8) with GWAS hits associated with protein C levels, triglycerides, serum urate levels, lipid metabolism, metabolic syndrome, and gamma-glutamyl transferase levels (S14 Table). The rs6976930 SNP shows an eQTL association with MLXIPL expression in omental (P = 7x10^-14) and subcutaneous adipose tissue (P = 4x10^-14). MLXIPL is 122 kb downstream of rs6976930 and codes for a transcription factor that binds carbohydrate response motifs, increasing transcription of genes involved in glycolysis, lipogenesis, and triglyceride synthesis [75, 76].</td>
<td><strong>0.8</strong></td>
<td>[75, 76]</td>
</tr>
<tr>
<td><strong>PLCE1 (rs10786152):</strong></td>
<td>There are 8 genes within 500 kb of the lead SNP (S3 Fig). The lead SNP lies within the intron of PLCE1 encoding a phospholipase involved in cellular growth and differentiation and gene expression among many other biological processes involving phospholipids [77]. Variants in this gene have been shown to cause nephrotic syndrome, type 3 [78]. Nearby variants rs9663362 and rs932764 (r^2 = 1.0 and 0.85, respectively) have been previously associated with systolic and diastolic blood pressure (S14 Table).</td>
<td><strong>0.8</strong></td>
<td>[77, 78]</td>
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<tr>
<td><strong>CTR2B (rs889512):</strong></td>
<td>The lead SNP showed an association with WHR_{adjBMI} in women only (Table 1). There are 17 genes within 500 kb (S3 Fig). The nearby rs4888378 SNP has been associated with carotid intima-media thickness in women but not in men, and BCA11 (breast cancer anti-estrogen resistance protein 1) has been implicated as the causal gene [79]. The rs4888378 SNP is not, however, in LD with our lead SNP (r^2&lt;0.1). The SNP rs7202877, in moderate LD with rs889512 (r^2 = 0.8), is a risk variant for type 1 diabetes (S14 Table). The data from Roadmap Epigenomics Consortium [22] suggest that five variants in strong LD (r^2&gt;0.8) with our lead SNP rest in known regulatory regions, including rs9393650 within an active enhancer region and rs72802352 in a DNAse hypersensitive region for human skeletal muscle cells and myoblasts; and rs147630228 and rs111869668 within active enhancer regions for the pancreas. Additionally, rs111869668 rests within binding motifs for CEBPB and CEBPD (CCATC enhancer-binding protein-Beta and Delta) which are enhancer proteins involved in adipogenesis [80, 81].</td>
<td><strong>0.8</strong></td>
<td>[79, 22, 80, 81]</td>
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</table>
explain less of the BMI variance among physically active compared to inactive individuals indicates that further interactions may exist, but larger meta-analyses, more accurate and precise measurement of PA, and/or improved analytical methods will be required to identify them. We found no difference between inactive and active individuals in variance explained by common SNPs in aggregate for WC_{adjBMI} or WHR_{adjBMI}, and no loci interacted with PA on WC_{adjBMI} or WHR_{adjBMI}. Therefore, PA may not modify genetic influences as strongly for body fat distribution as for overall adiposity. Furthermore, while differences in variance explained by common variants may be due to genetic effects being modified by PA, it is important to note that heritability can change in the absence of changes in genetic effects, if environmental variation differs between the inactive and active groups. Therefore, the lower BMI variance explained in the active group could be partly due to a potentially greater environmental variation in this group.

While we replicated the previously observed interaction between FTO and PA [7], it remains unclear what biological mechanisms underlie the attenuation in FTO’s effect in physically active individuals, and whether the interaction is due to PA or due to confounding by other environmental exposures. While some studies suggest that FTO may interact with diet [24–26], a recent meta-analysis of 177,330 individuals did not find interaction between FTO and dietary intakes of total energy, protein, carbohydrate or fat [27]. The obesity-associated FTO variants are located in a super-enhancer region [28] and have been associated with DNA methylation levels [29–31], suggesting that this region may be sensitive to epigenetic effects that could mediate the interaction between FTO and PA.

In genome-wide analyses for SNP main effects adjusting for PA, or when testing for the joint significance of SNP main effect and SNPxPA interaction, we identify 11 novel adiposity loci, even though our sample size was up to 40% smaller than in the largest published main effect meta-analyses [17, 18]. Our findings suggest that accounting for PA may facilitate the discovery of novel adiposity loci. Similarly, accounting for other environmental factors that contribute to variation in adiposity could lead to the discovery of additional loci.

In the present meta-analyses, statistical power to identify SNPxPA interactions may have been limited due to challenges relating to the measurement and statistical modeling of PA [5]. Of the 60 participating studies, 56 assessed PA by self-report while 4 used wearable PA monitors. Measurement error and bias inherent in self-report estimates of PA [32] can attenuate effect sizes for SNPxPA interaction effects towards the null [33]. Measurement using PA monitors provides more consistent results, but the monitors are not able to cover all types of activities and the measurement covers a limited time span compared to questionnaires [34]. As sample size requirements increase nonlinearly when effect sizes decrease, any factor that leads to a deflation in the observed interaction effect estimates may make their detection very difficult, even when very large population samples are available for analysis. Finally, because of the wide differences in PA assessment tools used among the participating studies, we treated PA as a dichotomous variable, harmonizing PA into inactive and active individuals. Considerable loss of power is anticipated when a continuous PA variable is dichotomized [35]. Our power could be enhanced by using a continuous PA variable if a few larger studies with equivalent, quantitative PA measurements were available.

In summary, while our results suggest that adjusting for PA or other environmental factors that contribute to variation in adiposity may increase power for gene discovery, we do not find evidence of SNPxPA interaction effects stronger than that observed for FTO. While other SNPxPA interaction effects on adiposity are likely to exist, combining many small studies with varying characteristics and PA assessment tools may be inefficient for identifying such effects [5]. Access to large cohorts with quantitative, equivalent PA variables, measured with relatively high accuracy and precision, may be necessary to uncover novel SNPxPA interactions.
Methods

Main analyses

Ethics statement. All studies were conducted according to the Declaration of Helsinki. The studies were approved by the local ethical review boards and all study participants provided written informed consent for the collection of samples and subsequent analyses.

Outcome traits—BMI, WC_{adjBMI} and WHR_{adjBMI}. We examined three anthropometric traits related to overall adiposity (BMI) or body fat distribution (WC_{adjBMI} and WHR_{adjBMI}) [36] that were available from a large number of studies. Before the association analyses, we calculated sex-specific residuals by adjusting for age, age^2, BMI (for WC_{adjBMI} and WHR_{adjBMI} traits only), and other necessary study-specific covariates, such as genotype-derived principal components. Subsequently, we normalized the distributions of sex-specific trait residuals using inverse normal transformation.

Physical activity. Physical activity was assessed and quantified in various ways in the participating studies of the meta-analysis (S1 and S6 Tables). Aiming to amass as large a sample size as possible, we harmonized PA by categorizing it into a simple dichotomous variable—physically inactive vs. active—that could be derived in a relatively consistent way in all participating studies, and that would be consistent with previous findings on gene-physical activity interactions and the relationship between activity levels and health outcomes. In studies with categorical PA data, individuals were defined inactive if they reported having a sedentary occupation and being sedentary during transport and leisure-time (<1 h of moderate intensity leisure-time or commuting PA per week). All other individuals were defined physically active. Previous studies in large-scale individual cohorts have demonstrated that the interaction between FTO, or a BMI-increasing genetic risk score, with physical activity, is most pronounced approximately at this activity level [6, 37, 38]. In studies with continuous PA data, PA variables were standardized by defining individuals belonging to the lowest sex- and age-adjusted quintile of PA levels as inactive, and all other individuals as active. The study-specific coding of the dichotomous PA variable in each study is described in S6 Table.

Study-specific association analyses. We included 42 studies with genome-wide data, 10 studies with Metabochip data, and eight studies with both genome-wide and Metabochip data. If both genome-wide and Metabochip data were available for the same individual, we only included the genome-wide data (S1 Table). Studies with genome-wide genotyped data used either Affymetrix or Illumina arrays (S2 Table). Following study-specific quality control measures, the genotype data were imputed using the HapMap phase II reference panel (S2 Table). Studies with Metabochip data used the custom Illumina HumanCardio-Metabo BeadChip containing ~195K SNPs designed to support large-scale follow-up of known associations with metabolic and cardiovascular traits [39]. Each study ran autosomal SNP association analyses with BMI, WC_{adjBMI} and WHR_{adjBMI} across their array of genetic data using the following linear regression models in men and women separately: 1) active individuals only; 2) inactive individuals only; and 3) active and inactive individuals combined, adjusting for the PA stratum. In studies that included families or closely related individuals, regression coefficients were estimated using a variance component model that modeled relatedness in men and women combined, with sex as a covariate, in addition to the sex-specific analyses. The additive genetic effect for each SNP and phenotype association was estimated using linear regression. For studies with a case-control design (S1 Table), cases and controls were analyzed separately.

Quality control of study-specific association results. All study-specific files for the three regression models listed above were processed through a standardized quality control protocol using the EasyQC software [40]. The study-specific quality control measures included checks on file completeness, range of test statistics, allele frequencies, trait transformation, population
stratification, and filtering out of low quality data. Checks on file completeness included screening for missing alleles, effect estimates, allele frequencies, and other missing data. Checks on range of test statistics included screening for invalid statistics such as P-values > 1 or < 0, negative standard errors, or SNPs with low minor allele count (MAC, calculated as MAF/N, where MAF is the minor allele frequency and N is the sample size) and where SNPs with MAC < 5 in the inactive or the active group were removed. The correctness of trait transformation to inverse normal was examined by plotting 2/median of the standard error with the square root of the sample size. Population stratification was examined by calculating the study specific genomic control inflation factor (λ_GC) [41]. If a study had λ_GC > 1.1, the study analyst was contacted and asked to revise the analyses by adjusting for principal components. The allele frequencies in each study were examined for strand issues and miscoded alleles by plotting effect allele frequencies against the corresponding allele frequencies from the HapMap2 reference panel. Finally, low quality data were filtered out by removing monomorphic SNPs, imputed SNPs with poor imputation quality (r²_hat < 0.3 in MACH [42], observed/expected dosage variance < 0.3 in BIMBAM [43], proper_info < 0.4 in IMPUTE [44]), and genotyped SNPs with a low call-rate (< 95%) or that were out of Hardy-Weinberg equilibrium (P < 10⁻⁶).

**Meta-analyses.** Beta-coefficients and standard errors were combined by an inverse-variance weighted fixed effect method, implemented using the METAL software [45]. We performed meta-analyses for each of the three models (active, inactive, active + inactive adjusted for PA) in men only, in women only, and in men and women combined. Study-specific GWAS results were corrected for genomic control using all SNPs. Study-specific Metabochip results as well as the meta-analysis results for GWAS and Metabochip combined were corrected for genomic control using 4,425 SNPs included on the Metabochip for replication of associations with QT-interval, a phenotype not correlated with BMI, WC_adjBMI or WHR_adjBMI, after pruning of SNPs within 500 kb of an anthropometry replication SNP. We excluded SNPs that 1) were not available in at least half of the maximum sample size in each stratum; 2) had a heterogeneity I² > 75%, or 3) were missing chromosomal and base position annotation in dbSNP.

**Calculation of the significance of SNP×PA interaction and of the joint significance of SNP main effect and SNP×PA interaction.** To identify SNP×PA interactions, we used the EasyStrata R package [46] to test for the difference in meta-analyzed beta-coefficients between the active and inactive groups for the association of each SNP with BMI, WC_adjBMI and WHR_adjBMI. Easystrata tests for differences in effect estimates between the active and inactive strata by subtracting one beta from the other (β_active - β_inactive) and dividing by the overall standard error of the difference as follows:

\[
Z_{diff} = \frac{\beta_{active} - \beta_{inactive}}{\sqrt{SE_{active}^2 + SE_{inactive}^2 - 2 \cdot r \cdot SE_{active}^2 \cdot SE_{inactive}^2}}
\]

where \(r\) is the Spearman rank correlation coefficient between \(\beta_{active}\) and \(\beta_{inactive}\) for all genome-wide SNPs. The joint significance of the SNP main and SNP×PA interaction effects was estimated using the method by Aschard et al. [16] which is a joint test for genetic main effects and gene-environment interaction effects where gene-environment interaction is calculated as the difference in effect estimates between two exposure strata, accounting for 2 degrees of freedom.

**Testing for secondary signals.** Approximate conditional analyses were conducted using GCTA version 1.24 [19]. In the analyses for SNPs identified in our meta-analyses of European-ancestry individuals only, LD correlations between SNPs were estimated using a reference
sample comprised of European-ancestry participants of the Atherosclerosis Risk in Communities (ARIC) study. In the analyses for SNPs identified in our meta-analyses of all ancestries combined, the reference sample comprised 93% of European-ancestry individuals and 6% of African ancestry participants from ARIC, as well as 1% of CHB and JPT samples from the HapMap2 panel, to approximate the ancestry mixture in our all ancestry meta-analyses. To test if our identified SNPs were independent secondary signals that fell within 1 Mbp of a previously established signal, we used the GCTA—cojo-cond command to condition our lead SNPs on each previously established SNP in the same locus.

Replication analysis for the **CDH12** locus. The replication analysis for the **CDH12** locus included participants from the EPIC-Norfolk (N\text{INACTIVE} = 4,755, N\text{ACTIVE} = 11,526) and Fenland studies (N\text{INACTIVE} = 1,213, N\text{ACTIVE} = 4,817), and from the random subcohort of the EPIC-InterAct Consortium (N\text{INACTIVE} = 2,154, N\text{ACTIVE} = 6,632). PA stratum-specific estimates of the association of **CDH12** with BMI were assessed and meta-analyzed by fixed effects meta-analyses, and the differences between the PA-strata were determined as described above.

Examining the influence of BMI, WC\text{adjBMI} and WHR\text{adjBMI}-associated loci on other complex traits and their potential functional roles

**NHGRI-EBI GWAS catalog lookups.** To identify associations of the novel BMI, WC\text{adjBMI} or WHR\text{adjBMI} loci with other complex traits in published GWAS, we extracted previously reported GWAS associations within 500 kb and \(r^2 > 0.6\) with any of the lead SNPs, from the GWAS Catalog of the National Human Genome Research Institute and European Bioinformatics Institute [47] (S14 Table).

**eQTLs.** We examined the ***cis***-associations of the novel BMI, WC\text{adjBMI} or WHR\text{adjBMI} loci with the expression of nearby genes from various tissues by performing a look-up in a library of >100 published expression datasets, as described previously by Zhang et al [48]. In addition, we examined ***cis***-associations using gene expression data derived from fasting peripheral whole blood in the Framingham Heart Study [49] (n = 5,206), adjusting for PA, age, age\(^2\), sex and cohort. For each novel locus, we evaluated the association of all transcripts \(\pm 1\) Mb from the lead SNP. To minimize the potential for false positives, we only considered associations where our lead SNP or its proxy (\(r^2 > 0.8\)) was either the peak SNP associated with the expression of a gene transcript in the region, or in strong LD (\(r^2 > 0.8\)) with the peak SNP.

**Overlap with functional regulatory elements.** We used the Uncovering Enrichment Through Simulation method to combine the genetic association data with the Roadmap Epigenomics Project segmentation data [22]. First, 10,000 sets of random SNPs were selected among HapMap2 SNPs with a MAF > 0.05 that matched the original input SNPs based on proximity to a transcription start site and the number of LD partners (\(r^2 > 0.8\) in individuals of European ancestry in the 1000 Genomes Project). The LD partners were combined with their original lead SNPs to create 10,000 sets of matched random SNPs and their respective LD partners. These sets were intersected with the 15-state ChromHMM data from the Roadmap Epigenomics Project and resultant co-localizations were collapsed from total SNPs down to loci, which were then used to calculate an empirical P value when comparing the original SNPs to the random sets. We examined the enrichment for all loci reaching P < 10\(^{-5}\) for SNP×PA interaction combined, and for all loci reaching P < 5x10\(^{-8}\) in the PA-adjusted SNP main effect model combined. In addition, we examined the variant-specific overlap with regulatory elements for each of the index SNPs of the novel BMI, WC\text{adjBMI} and WHR\text{adjBMI} loci and variants in strong LD (\(r^2 > 0.8\)).

**Estimation of variance explained in inactive and active groups.** We compared variance explained for BMI, WC\text{adjBMI} and WHR\text{adjBMI} between the active and inactive groups using
two approaches. First, we used a method previously reported by Kutalik et al [15], and selected subsets of SNPs based on varying P value thresholds (ranging from $5 \times 10^{-8}$ to 0.05) from the SNP main effect model adjusted for PA. Each subset of SNPs was clumped into independent regions using a physical distance criterion of < 500kb, and the most significant lead SNP within the respective region was selected. For each lead SNP, the explained variance was calculated as:

$$r^2 = \frac{1}{1 + \frac{N}{(1 + (\frac{N}{2} - 1)^2)}} - \frac{1}{N}$$

in the active and inactive groups separately, where $N$ is the sample size and $P$ is the P value for SNP main effect in active or inactive strata. Finally, the variance explained by each subset of SNPs in the active and inactive strata was estimated by summing up the variance explained by the SNPs.

Second, we applied the LD Score regression tool developed by Bulik-Sullivan et al [14] to quantify the proportion of inflation due to polygenicity (heritability) rather than confounding (cryptic relatedness or population stratification) using meta-analysis summary results. LD Score regression leverages LD between causal and index variants to distinguish true signals by regressing meta-analysis summary results on an ‘LD Score’, i.e. the cumulative genetic variation that an index SNP tags. To obtain heritability estimates by PA strata, we regressed our summary results from the genome-wide meta-analyses of BMI, WC_{adjBMI} and WHR_{adjBMI} stratified by PA status (active and inactive), on pre-calculated LD Scores available in HapMap3 reference samples of up to 1,061,094 variants with MAF $\geq$ 1% and $N > 10^{th}$ percentile of the total sample size.

Supporting information

S1 Acknowledgements. A full list of acknowledgements.

S1 Fig. Interaction between the $CDH12$ locus and physical activity on BMI in the discovery genome-wide meta-analysis (n = 134,767), in the independent replication sample (n = 31,097), and in the discovery and replication samples combined.

S2 Fig. Quantile-Quantile and Manhattan plots for the genome-wide meta-analysis results of the SNP main effect adjusting for physical activity (SNPadjPA), interaction between SNP and physical activity, and the joint effect of SNP main effect and SNP × PA interaction (Joint2df) in men and women of European-ancestry combined.

S3 Fig. Regional association plots for novel BMI, WC_{adjBMI} or WHR_{adjBMI} loci showing either a genome-wide significant SNP main effect when adjusting for physical activity as a covariate, or a genome-wide significant joint effect of physical activity-adjusted SNP main effect and SNP × physical activity interaction.

S4 Fig. Heatmap of P values for the physical activity-adjusted SNP main effect model (PadjPA), the joint model (Pjoint), and the SNP x PA interaction model (Pint).
S1 Table. Basic study information and description of outcome assessment (BMI, WC, WHR) and Physical activity assessment.
(XLSX)

S2 Table. Genotyping and imputation platforms of the participating studies.
(XLSX)

S3 Table. Population characteristics for inactive and active individuals combined in the participating studies.
(XLSX)

S4 Table. Population characteristics for inactive individuals in the participating studies.
(XLSX)

S5 Table. Population characteristics for active individuals in the participating studies.
(XLSX)

S6 Table. Methods used for measuring physical activity and definitions of inactive for studies participating in the meta-analyses.
(XLSX)

S7 Table. All SNPs that met significance for BMI in the European only analyses for at least one of the approaches tested: interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S8 Table. All SNPs that met significance for BMI in the all ancestry analyses for at least one of the approaches tested: interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S9 Table. All SNPs that met significance for waist circumference adjusted for BMI in the European only analyses for at least one of the approaches tested: interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S10 Table. All SNPs that met significance for waist circumference adjusted for BMI in the all ancestry analyses for at least one of the approaches tested: interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S11 Table. All SNPs that met significance for waist-to-hip ratio adjusted for BMI in the European only analyses for at least one of the approaches tested: interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S12 Table. All SNPs that met significance for waist-to-hip ratio adjusted for BMI in the all ancestry analyses for at least one of the approaches tested: interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
(XLSX)

S13 Table. Variance explained using P value thresholds.
(XLSX)

S14 Table. GWAS catalog lookups for novel loci and new secondary signal in known loci.
(XLSX)
S15 Table. Association of the novel loci with cis gene expression (cis-eQTL).
(XLSX)

S16 Table. Association of loci identified for interaction with physical activity, for physical activity-adjusted SNP main effect, or for joint association of SNP main effect and physical activity interaction, with physical activity and sedentary behaviour.
(XLSX)

S17 Table. Results for approximate conditional analyses to identify secondary signals in the novel BMI, WC_{adjBMI} or WHR_{adjBMI}-associated loci.
(XLSX)

S18 Table. Enrichment of loci interacting with PA (P_{int}<10^{-5}) on the level of BMI with functional genomic elements in adipose, brain, and muscle tissue cell lines from the Roadmap Epigenomics Project.
(XLSX)

S19 Table. Enrichment of loci interacting with PA (P_{int}<10^{-5}) on the level of WHR_{adjBMI} with functional genomic elements in adipose, brain, and muscle tissue cell lines from the Roadmap Epigenomics Project.
(XLSX)

S20 Table. Enrichment of loci showing association with BMI (P_{adjPA}<5x10^{-8}) with functional genomic elements in adipose, brain, and muscle tissue cell lines from the Roadmap Epigenomics Project.
(XLSX)

S21 Table. Enrichment of loci showing association with WHR_{adjBMI} (P_{adjPA}<5x10^{-8}) with functional genomic elements in adipose, brain, and muscle tissue cell lines from the Roadmap Epigenomics Project.
(XLSX)

S22 Table. Association of novel loci identified for interaction with physical activity, for physical activity-adjusted SNP main effect, or for joint association of the SNP main effect and physical activity interaction, in GIANT results not accounting for physical activity.
(XLSX)

**Author Contributions**

**Conceptualization:** TOK RJFL.

**Data curation:** TOK LAC RJFL KLMon.

**Formal analysis:** RAS KLY AEJ TWW AMa DHa NLHC JSN TSA LQ TW FRe MdH TOK QQ MG TSA LX AYC LAC MC RJFL MFF KEN ADJ JEHa RJK.

**Investigation:** AVS TBH GE LJJ VG KEN MG AJ KLMon KLY EB PGL JBW NGM GWM DPS GC LJP JHua AW M AlJ JB MFo CBe HMS TR SSn BS YW JBB LSA ZK PMMV TC Sbi GWi PV KKf ASH KHa SM VS MP JM IR CHeup VV IK OPo LY DT BB MMMar AB PF RRaw TAL PK MfHo KSa RMag THea TEB Ama JL RAS SBr NJW JHZ RL SBu AD NA CMvD IBB MFF LB RJFL LX NLHC EL JP PJG CSF CTL LAC MB FSC KLMol RNB JT LL TEs HP CS HAK LFB SLRK MAJ PAP SA FRe IB GH PWF JE CO MLo AEJ JOJ TSA LPa GDS TIAS JEHu DJP SP LJJ BHS SSa YCC MFu JRO ARS MHa MA AL HVe LQu THu QQ JAS JDF WZhan DRW KHv OLH AUJ LLB AJS KKv TTan DHe SBa MNH MJM KF
NG OPe TWW IMH MO CG HG AP KS t CHo C Hut RRau BT MMN WZhao BL WRS STT JCC JSDK MEG GED TBG GS JHui WM UL CHa LL LH KL AGB LJRT JAN YL MLa JK AS PSC NN WJP GvG YM BWJHP AMu CML MIM TTam JA MRJ LQi THan DK KKO AW AJ UG EJcdG MHMdM JJH DIB GWA PN AFN NDH SW HC JFW CBo UV MD LPe DP MW TMF NVR MCZ FRE AH AGU JLB G SS FB AMe GRA FC ATE HvO UV MD SG MN RMan IP ATo PJvdM JvVO IMH HS AJO CAH MMAN DIC AYC LMR PMR SBI NZ JG UP CK MKu MK CL LPL NHK MJ MKa OTR TL.

**Supervision:** GRA IB MB IBB CSF TMF IMH RJFL MIM KLMon KEN JRO DPS CMvD JNH.

**Writing – original draft:** MG RAS AEJ KLY MFF LB LQu PWF RJFL TOK.

**Writing – review & editing:** MG RAS AEJ KLY MFF TWW LB AYC AMa DHa LX TW NLHC TSA MDH QQ JSN FRe LQu JDE JEHA MC ZK EL JL JHEU WZh JZhan WZhao P JG T Hal SA PMMV SBI LY ATe AVS MKu MNH JM JMEK M Ho NVR JBJH JHZ HS LL Chup GWi WP JW KKr AD KL YL M Fo MHa LB GC TTa n RMa g PJvdM AUJ JLBG VV JM PN CBe DP AJ SSn YCC JE UL MA LSA NA BB JA JN RNB AGB JB AB LLL JBB SBr FB SBu HC PSC FSC TC GDS GED ND MD TEB GE TEs JDF MFu KF CG SJG JG PGL HG TNG NO GvG ThU KhA NDH AS DHe LH LJH OLH Cho JJH Jhua THan HJui ChUt NHH ALJ JOJ MAH MJ LH AK IK PK JK KKv MKa TAL LJL B CL MLo RL MMa r YM KLMon GWM MHHdM A Mu AMe RMan SM NN MN JAIN IMN AJO MO KKO SP LPa JP MP AP UP PAP IP HP OTR TR LJRT RRau PMR LMR IR CS MAS Ksa WR SSSa ARS SSI GS BHS JAS HS AS BS AIS TTa n STT BT DT LV HVe JvVO MCV UV GWA MW SW AW AFW MCZ NZ C AHai LL UG CO AH FrI AGU LPe Fw CHa OPo FC KhV CAHar ATo SBA LJ LSP SLRK RRau TIAS JT VS BWJHP EJcdG DIB TL MMa n MLa CBo NGM DK AL WM KSt MKi TBH VG HVo LQI MRJ JCC PK CF CK PV GH OPe NJW CL DRW DJP EB DIC GRA IB MIM TMF JRO CMvD MB IMH KLMoL DPS CSF CTL JNH RJK ADJ IBJ PWF KEN LAC RJFL TOK.

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Correction: Genome-wide physical activity interactions in adiposity — A meta-analysis of 200,452 adults

There are errors in some of the gene names in S7, S8, S9, S10, S11 and S12 Tables. Please view the correct tables below, with the locus numbers removed since they are not defined across all the models in the paper.

**Supporting information**

**S7 Table.** All SNPs that met significance for BMI in the European only analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.

**S8 Table.** All SNPs that met significance for BMI in the all ancestry analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.

**S9 Table.** All SNPs that met significance for waist circumference adjusted for BMI in the European only analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.

**S10 Table.** All SNPs that met significance for waist circumference adjusted for BMI in the all ancestry analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.

**S11 Table.** All SNPs that met significance for waist-to-hip ratio adjusted for BMI in the European only analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.

**S12 Table.** All SNPs that met significance for waist-to-hip ratio adjusted for BMI in the all ancestry analyses for at least one of the approaches tested: Interaction, adjusted for physical activity, or jointly accounting for the main and interaction effects.
Reference